



Hancock Stain User Guide

Introduction:

Hancock Stain (P/N HCS-101) is an Eosin-Nigrosin stain used to determine the number of live-dead spermatozoa in an ejaculate and to analyze morphological characteristics. Spermatozoa with intact cell membranes will not absorb the stain while those with damaged cell membranes will absorb the stain and appear darker.

This stain is used as an aid for reproductive evaluation. More information on the morphological examination of equine spermatozoa can be found in the book Equine Reproduction by McKinnon and Voss and/or the Colorado State University Bulletin #5, Management of the Stallion for Maximum Reproductive Efficiency, II.

Slide Preparation:

The following instructions show the steps necessary to prepare a slide for morphological evaluation.

1. In order to prevent morphological abnormalities due to cold shock, pre-warm a slide to 37°C using a slide warmer (P/N 581-MOD1). Once slide is warm, place 1 drop of Hancock Stain solution on one end of the slide (Figure 1).

CAUTION: Placing semen on the slide before the stain will contaminate the Hancock Stain.

2. Using a disposable pasteur pipette (P/N 537-503), place 1 drop of fresh semen next to the stain (Figure 2) and use the tip of the pipette to gently mix (Figure 3).



Figure 1.
Placement of stain
on slide.



Figure 2.
Placement of
semen on slide.

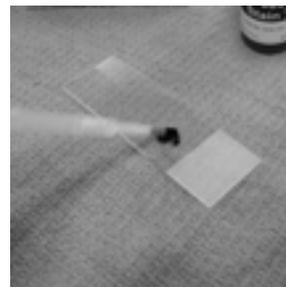


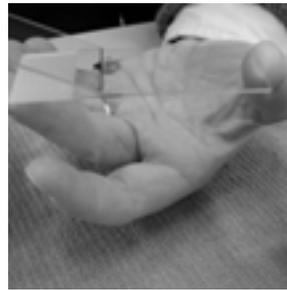
Figure 3.
Mixing stain and
semen.

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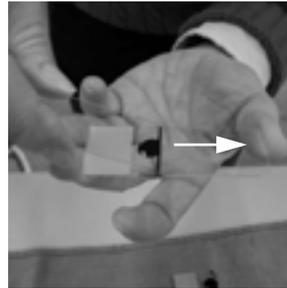
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3. Use the edge of another slide, held at a 45 degree angle (Figure 4), to touch the mixture and allow the stain to collect along the bottom edge of the second slide. Push the second slide forward to create a smear (Figure 5).
4. Quickly dry the slide by waving it back and forth, setting it on a slide warmer, or using a hair dryer.
5. Using a 400x or greater magnification microscope, observe the sperm cells located in the feathered region of the slide at the edge furthest from where the initial droplets were deposited (Figure 6).
6. Determine the percent live cells by counting 100 cells throughout the slide for each of two slides and recording the average number of live (unstained) versus dead (stained) cells.
7. Determine the percent normal cells by counting 100 cells throughout the slide for each of two slides and recording the average number of normal versus abnormal sperm observed.



*Figure 4.
Using another
slide to spread
mixture.*



*Figure 5.
Push second slide
forward to create
smear.*



*Figure 6.
View feathered
edge under
microscope.*

Storage:

Store bottle in a cool location.

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